

Cell Types and Events in Graft Infiltrates that Lead to Acceptance or Rejection of Cardiac Allografts in Neonatally-Tolerized Mice V. Giannitsos, R.A. Bascom, K. Tao and L.J. West

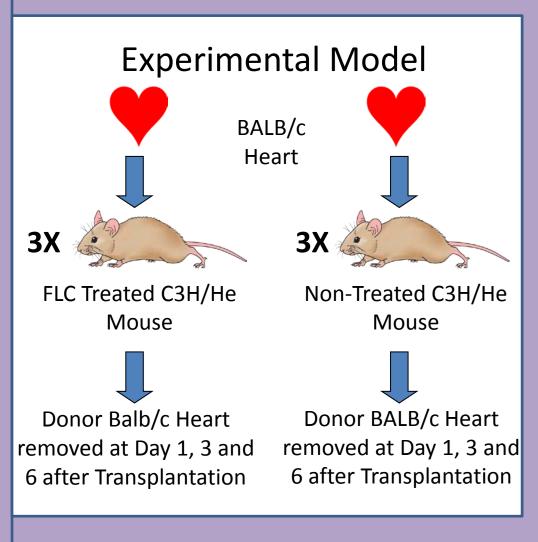
Department of Pediatrics, Surgery and Immunology, University of Alberta, Edmonton, Canada

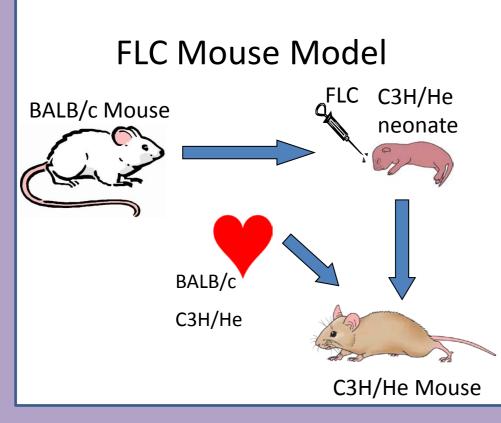
Introduction

Acute rejection of an organ graft is often associated with infiltration by T cells and other cell types, causing damage to the blood vessels and tissues. Long-term acceptance of organ grafts without immunosuppressive drug therapies is a major goal of transplantation research. Many strategies have been explored to induce transplant tolerance in animal models. One strategy to induce cardiac allograft acceptance involves injecting neonatal C3H/He mice $(H-2^k)$ with allogeneic fetal liver cells (FLC) from BALB/c mice $(H-2^d)$. Typically, this induces acceptance of BALB/c cardiac grafts in >90% of recipients, while 100% of untreated mice reject grafts. Our lab is currently studying the mechanisms of graft acceptance in this model. The objective of this project was to investigate how the cell types and events in graft infiltrates differ between FLC-treated and untreated mice, in an attempt to understand the processes leading to cardiac graft acceptance.

Methods

Fetal livers were collected from BALB/c mice (H-2^d) at approximately 15 days gestation and made into a single cell suspension for injection. Three neonatal C3H/He mice (H-2^k) BALB/c Mouse were injected with FLC within 24 hours of birth and three other C3H/He neonates were left untreated. At approximately six weeks, the mice heterotopic heart underwent transplantation from BALB/c donors. Two mice were sacrificed one day after transplantation: one FLC- treated and one untreated.





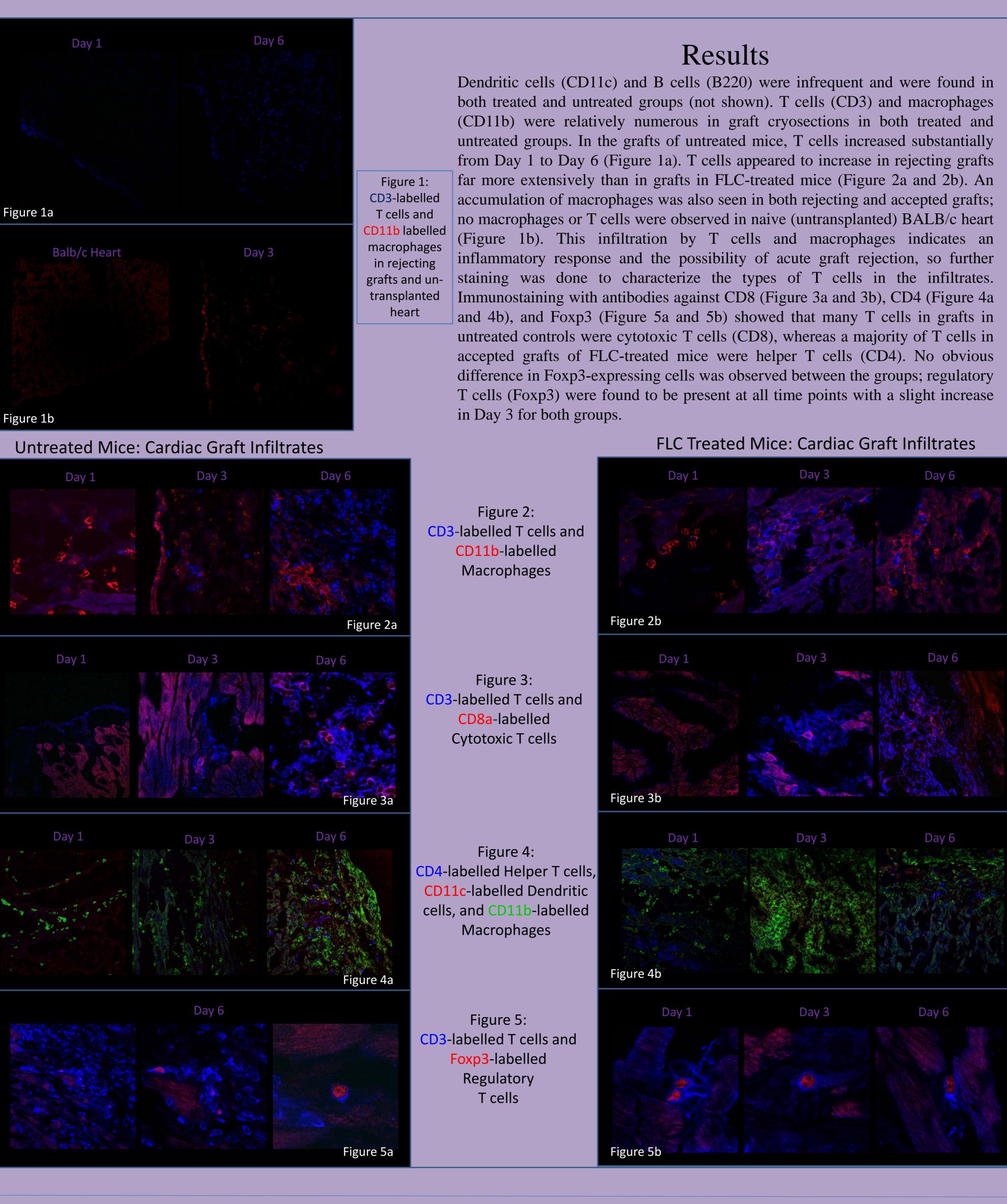
The BALB/c donor and C3H/He native hearts were collected along with other C3H/He organs. This process was repeated at days 3 and 6 post-transplant for the remaining mice. The six donor hearts and a naive BALB/c control heart were made into cryosections. The cryosections were labelled either directly or indirectly with antibodies to identify different cell types by confocal microscopy.

Antibody Chart			
Target	Label	lsotype	Clone
CD3 (T Cells)	Alexa Fluor 647	Rat(SD) IgG _{2b} , κ	17A2
CD4 (Helper T Cells)	Alexa Fluor 647	Rat(DA) IgG _{2a} , κ	RM4-5
CD8a (Cytotoxic T Cells)	Biotin	Rat IgG _{2a} , κ	53-6.7
CD11b (Macrophages)	Biotin	Rat IgG _{2b} , κ	M1/70
CD11c (Dendritic Cells)	Purified	Ar Ham lgG ₁ , λ_2	HL3
Foxp3 (Regulatory T Cells)	Biotin	Rat IgG _{2a} , κ	FJK-16s
CD45R (B Cells)	Biotin	Rat IgG _{2a} , κ	RA3-6B2

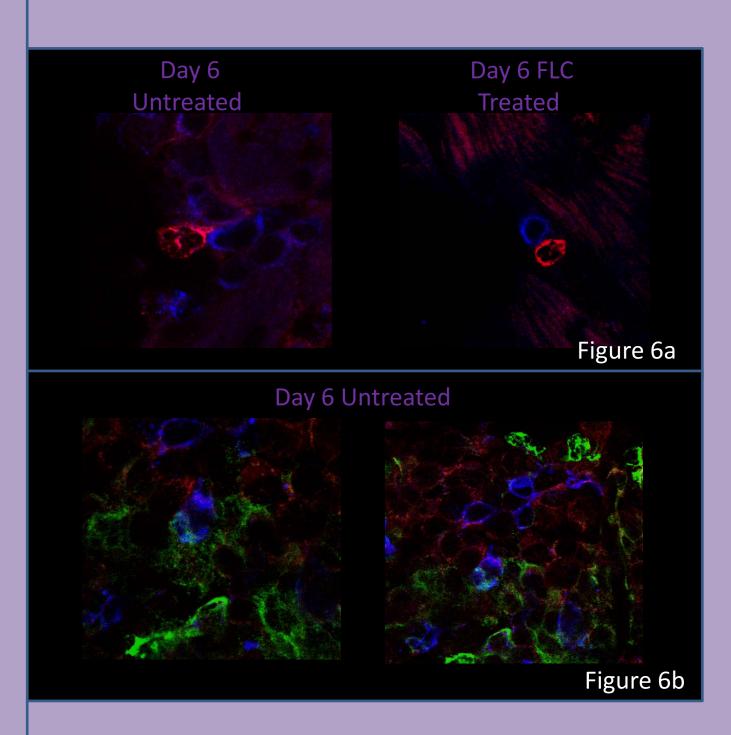
A satile a des Clease

Acknowledgments

I would like to thank AHFMR for the funding and allowing me this opportunity to work in research. As well I would like to thank Dr. Lori West for guidance and giving me the chance to work in her lab. Thanks are given to Tao for doing the heart transplants and Roger for helping me with the microscope and all other aspects of my project. Lastly thank you to the entire West Lab Group for making my summer enjoyable. The West Research Group is funded by: Heart and Stroke Foundation of Canada, AHFMR, CIHR, NIH(USA), NSERC/CIHR(CHRP), CRC/CFI, ISHLT, Stollery Children's Hospital Foundation, WCHRI (Women and Children's Health Research Institute).



Other results observed in this qualitative experiment were the apparent cellto-cell interactions between T cells and macrophages in both grafts from FLC-treated and untreated mice (Figure 6a). In a small number of cases, interactions occurred between helper T cells (CD4) and dendritic cells in Day 6 of the untreated control (Figure 6b).



Infiltration of allografts by T cells and macrophages indicates an inflammatory and/or immune response (ie, acute allograft rejection). This T cell-mediated response was observed in both grafts from FLC-treated and untreated mice but the extent to which the T cells increased in accepted grafts was less than in the rejecting grafts. Infiltrating T cells in the grafts of FLC-treated mice could probably be attributed to the alloantigenindependent response of ischemia-reperfusion caused during transplant surgery. In addition, in the untreated mice alloantigen-dependant events can stimulate rejection of the allograft and tissue damage through the activation of donor-specific cytotoxic and helper T cells. Qualitative assessment of graft infiltrating cells suggests less cytotoxic T cells in FLC-treated recipients when compared to rejecting grafts in untreated controls. Further analysis of cellular interactions and events in the graft infiltrates may provide further information on the mechanisms of graft acceptance in this model.

Future Research

•Examine damage to the blood vessels and tissue by immunostaining with antibodies against active Caspase-3 and Complement. •Analyze the cell types and interactions of FLC-Treated Hearts surviving past 6 days.

•Examine spleen and lymph nodes collected from FLC-treated and untreated mice for cell types and interactions. •Determine if there is a difference in number of alloreactive regulatory T cells in the graft infiltrates of FLC-treated and untreated mice.

Further Information

Please contact giannits@ualberta.ca. More information on this and related projects can also be obtained at the West Research group website http://www.cardiactransplantresearch.med.ualberta.ca/index.php.



Results (continued)

Figure 6: CD4-labelled Helper T cells, CD11c-labelled Dendritic cells, and **CD11b**-labelled Macrophages

Conclusion