THE ROLE OF REGULATORY T CELLS IN THE DEVELOPMENT OF B-CELL TOLERANCE AFTER ABO-INCOMPATIBLE HEART TRANSPLANTION

Malika Ladha, Dr. Esmé Dijke, Dr. Lori West. Departments of Pediatrics, Surgery, and Medical Microbiology and Immunology, University of Alberta, Edmonton, Alberta.

INTRODUCTION

In adult heart transplantation, blood-group incompatibility between donors and recipients is a major barrier. ABOincompatible heart transplantation in infants, however, leads to the development of tolerance to donor blood-group antigens. The mechanisms that mediate the development of B-cell tolerance to blood-group antigens are not known.

We hypothesize that regulatory CD4+CD25highFoxp3+ (Treg) cells are capable of inhibiting the proliferation and function of bloodgroup A/B-antigen-specific B cells by selectively killing these cells.

The aim of this study is to investigate the effects of Treg cells on A/B-antigen-specific B cells.



This study is currently being conducted by in vitro stimulation of B cells in the presence and absence of Tregs. The cells are isolated from peripheral blood samples from healthy individuals. We use flow cytometry to characterize the cells and ELISAs to determine the cell proliferation and production of total IgM and A/B-specific antibodies. In order to define the optimal protocol for the analysis, we initially: 1) compared two different isolation techniques for B-cells (i.e., magnetically-activated cell sorter (AutoMACS) and immunomagnet cell separator (EasySep)], 2) optimized the ELISA protocol for detecting bloodgroup antibodies in culture supernatant and 3) tested various culture conditions for B cells and T cells (round bottom vs. flat bottom; CpG vs. CpG + II-2 vs. CpG + II-2 + II-4; with or without serum [human or bovine]).



Four samples of PBMCs were divided into two groups each: one group underwent separation via a magnetically activated cell sorter, AutoMACS, while the other group underwent separation using an immunomagnet cell separator, EasySep. The populations were then analyzed for recovery, viability and purity. No differences were detected between EasySep and AutoMACS. The AutoMACS will be used in our



This experiment was conducted with blood type-A plasma and blood type-B plasma (data for type-B is not

accurate analysis of the binding of the antibodies to B antigens. The Super Blocking Blocking Buffer, on the

other hand, does not reduce the background. Also, this experiment demonstrated that the ideal coating

Isolation Techniques - Comparing the AutoMACS and EasySep, there were no differences in recovery, purity, viability and function of the B cells. ELISA -

Blocking with goat serum dramatically reduced the background. Other

changes which enabled us to more accurately determine the presence of

antibodies included increasing the number of washing steps and using a

solution with 5% goat serum as a diluent. Also, a difference in using frozen

and fresh supernatant was noted with the latter allowing for detection of

higher antibody levels (increased sensitivity). Culture Conditions - For

isolated B cells, the highest proliferation was detected after culturing in 96-

well flat bottom plates in AIM-V medium supplemented with CpG.

Interleukin (IL)-2 and IL-4. These conditions also yielded the highest levels of

shown). The blocking buffer with Goat Serum drastically reduced the background which enabled the

0.10

0.00

Figures 4-5: Optimizing ELISA for Antibody Production

concentration for the coating antibody is 5.0 ug/mL.

SUMMARY OF RESULTS:

antibody production.

0.40

0.20



FIGURES & RESULTS continued

CULTURE CONDITIONS:

Figure 7: Antibody ELISA of rnatant from Various Culture Conditions

proliferation ELISA was

An antibody ELISA was conducted on supernatant from the experiments testing various culture conditions. This particula experiment was conducted on PBMCs from a blood type-A sample. AIM-V supplemented with II-2 and II -4 also had the hight levels of antibody production

SUMMARY & FUTURE DIRECTIONS

For our Treg and B-cell study, we will isolate B cells and Treg cells by autoMACS and culture the cells using the following culture conditions: 96-well flat bottom plates in AIM-V medium supplemented with CpG, Interleukin (IL)-2 and IL-4. In addition, we will use agat serum as a component in the blocking and diluent solutions for the ELISA. The ELISA will be carried out with fresh supernatant from the in vitro stimulation of B cells in the presence and absence of Treas.

ACKNOWLEDGEMENTS Dr. Lori West Dr. Esmé Dijke Bruce Motvka WEST Lab Alberta Innovates Health Solutions